curio ketone 8c as predominantly one diastereomer. Oxymercuration of (E)-5c gave, as expected, predominantly the diastereomeric  $\alpha$ -mercurio ketone epi-8c; in both cases, the diastereoselectivity of the oxymercuration reaction approaches 90%. The absolute configuration at  $C_6$  of these intermediates remains to be established. Under normal Ferrier reaction conditions, (Z)-5c and (E)-5c afford the same ratio of inosose diastereomers, indicating that the configuration at the mercury-bearing carbon of 8c has no influence on the stereochemical outcome of the aldol reaction.

Although 8c appears to be remarkably stable as a dilute solution in aprotic solvents, exposure to Lewis acids results in rapid conversion to the inososes 7c, 12c, 13c, and 14c. Exploratory experiments indicate that the product ratio is highly dependent upon the Lewis acid promoter (Table II). Interestingly, the SnCl<sub>4</sub>-promoted cyclization of 8c is significantly more stereoselective than the cyclization of epi-8c. Clearly, the Lewis acid promoted version of the Ferrier reaction offers new possibilities for stereochemical control in intramolecular aldol reactions, and further characterization of this process is underway. In addition, application of this methodology to the total synthesis of biologically interesting inositol polyphosphates is in progress and will be reported in subsequent publications.<sup>16</sup>

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Supplementary Material Available: Experimental details and characterization data for new compounds 5a-d, 7a-d, 8c, 9c,d, 10a-d, 11b,c, 12a-d, 13c, and 14c (12 pages). Ordering information is given on any current masthead page.

(15) In some experiments, an initial oxymercuration adduct 15 (corresponding to the first intermediate in Scheme II) was obtained as the initial product, which, upon standing, lost MeOH at a variable rate to give 8c.

(16) One application of this methodology may be found in the accompanying paper: Estevez, V. A.; Prestwich, G. D. J. Am. Chem. Soc., following paper in this issue.

## Synthesis of Enantiomerically Pure, P-1-Tethered Inositol Tetrakis(phosphate) Affinity Labels via a Ferrier Rearrangement

Virginia A. Estevez and Glenn D. Prestwich\*

Department of Chemistry, State University of New York Stony Brook, New York 11794-3400

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D-myo-Inositol 1,4,5-tris(phosphate) (IP<sub>3</sub>) (1) (Figure 1) is an intracellular second messenger that mediates the release of calcium from nonmitochondrial stores¹ via binding to a transmembrane receptor protein.² Several other inositol polyphosphates have also been implicated in the regulation of calcium levels.³ Of these, D-myo-inositol 1,3,4,5-tetrakis(phosphate) (IP<sub>4</sub>) (2) may control regulation of Ca²+ reentry into the cell and modulate the IP<sub>3</sub>-sensitive Ca²+ pools.³ Clarification of the physiological role of IP<sub>4</sub> would be facilitated by the isolation and characterization of its cellular receptor (IP<sub>4</sub>R). Recently, we reported the synthesis⁴

Figure 1. D-myo-Inositol I,4,5-tris(phosphate) (IP<sub>3</sub>, 1), D-myo-inositol 1,3,4,5-tetrakis(phosphate) (IP<sub>4</sub>, 2), and derivation of P-1-modified IP<sub>4</sub> from the glucose carbon skeleton.

### Scheme Ia

<sup>a</sup>Reagents and conditions: (a) Trityl chloride, DAP, Et<sub>3</sub>N, DMF, room temperature, 12 h; (b) NaH, PMB-Cl, DMF, reflux, 12 h; (c) 5% H<sub>2</sub>SO<sub>4</sub>-MeOH, acetone, room temperature, 30 min; (d) oxalyl chloride, DMSO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, −78 °C to room temperature; (e) Ac<sub>2</sub>O, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN, 80 °C, 8 h; (f) (i) Hg(OAc)<sub>2</sub>, 3:2 acetone: water, room temperature, 30 min; (ii) saturated NaCl, room temperature, 24 h; (g) NaBH(OAc)<sub>3</sub>, HOAc, CH<sub>3</sub>CN, room temperature, 30 min; (h) BOM-Cl, Bu<sub>4</sub>NBr, H<sup>+</sup> sponge, CH<sub>3</sub>CN, room temperature, to 35 °C, to 55 °C; (i) NaOH, MeOH, reflux, 2 h; (j) (i) (i-Pr<sub>2</sub>N)-(OBn)P(OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>NHCbz), tetrazole, CH<sub>2</sub>Cl<sub>2</sub>, room temperature, 3 h; (ii) MCPBA, −48 °C for 3 min, 0 °C for 15 min; (k) DDQ, wet CH<sub>2</sub>Cl<sub>2</sub>, room temperature, 6 h; (l) (i) (BnO)<sub>2</sub>P(i-Pr<sub>2</sub>N), tetrazole, CH<sub>2</sub>Cl<sub>2</sub>, room temperature, 12 h; (ii) MCPBA, −48 °C to room temperature, 2 h; (m) (i) Pd-C, H<sub>2</sub>, 95% EtOH, 50 psi, room temperature, 5.5 h; (ii) Na-Chelex chromatography; (n) (i) NHS-ASA, DMF, 0.25 M TEAB (pH 8.0), room temperature, 12 h; (ii) DEAE cellulose chromatography; (iii) [<sup>125</sup>]Nal, lodobeads, 100 mM Na<sub>2</sub>HPO<sub>4</sub> (pH 7.5), room temperature, 10 min.

of a P-1-tethered<sup>5</sup> derivative of racemic IP<sub>4</sub> (3) and the corresponding bioaffinity matrix which allowed isolation and purifi-

<sup>(1) (</sup>a) Berridge, M. J. Annu. Rev. Biochem. 1987, 56, 159-193. (b) Berridge, M. J.; Irvine, R. F. Nature 1989, 341, 197-203.

<sup>(2)</sup> Supattapone, S.; Worley, P. F.; Baraban, J. M.; Snyder, S. H. J. Biol. Chem. 1988, 263, 1530-1534.

<sup>(3) (</sup>a) Nicoletti, F.; Bruno, V.; Fiore, L.; Cavallaro, S.; Canonico, P. J. Neurochem. 1989, 53, 1026–1030. (b) Vallejo, M.; Jackson, T.; Lightman, S.; Handley, M. Nature 1989, 330, 656–658. (c) Irvine, R. F.; Moore, R. M. Biochem. J. 1986, 240, 917–920.

<sup>(4)</sup> Estevez, V. A.; Prestwich, G. D. Tetrahedron Lett. 1991, 32, 1623-1626.

cation of rat brain IP<sub>4</sub>R.<sup>6</sup> The application of a radioiodinated photoaffinity analogue, [<sup>125</sup>I]ASA-IP<sub>4</sub>, for identification of specific binding subunits has also been described (ASA = 4-azido-salicylamide).<sup>7</sup> However, since the receptor only recognizes the D isomer of IP<sub>4</sub>,<sup>8</sup> we now describe a novel route to enantiomerically pure,<sup>9</sup> P-1-tethered IP<sub>4</sub> via a Ferrier rearrangement<sup>10</sup> of a suitably protected D-glucose derivative.

Selective modification required a protecting-group strategy that would differentiate the C-1 hydroxyl from the other three hydroxyls destined for phosphorylation. The stereochemical requirements of myo-inositol and the disposition of the phosphates in IP4 can be readily satisfied by a Ferrier rearrangement of a selectively protected D-glucose derivative11 (Figure 1). The synthesis of the optically active C-1 phosphodiester analogue of IP<sub>4</sub> bearing a reactive aminopropyl tether is summarized in Scheme I. Tritylation of methyl  $\alpha$ -D-glucopyranoside<sup>12</sup> followed by treatment with NaH and p-methoxybenzyl chloride provided the fully protected pyranoside in 63% overall yield. Selective detritylation was achieved in 90% yield with 5% H2SO4-MeOH and acetone as a cosolvent.13 Swern oxidation of alcohol 4 provided the aldehyde (>90% hydrated), and treatment with Ac<sub>2</sub>O and K2CO3 gave the (Z)-enol acetate 5 in 85% yield from 4.14 Ferrier rearrangement of 5 in the presence of mercuric trifluoroacetate and aqueous NaCl furnished the inosose 6 with the desired axial stereochemistry at the 2-OH (less than 10% equatorial)<sup>10</sup> in 60% yield. Stereoselective reduction of the  $\beta$ -hydroxy ketone with sodium triacetoxyborohydride15 provided the myoinositol skeleton 7 with the C-1 position differentially protected from the C-3, C-4, and C-5 hydroxyls.

We then focused on protecting the two remaining hydroxyls in diol 7. For racemic, P-1-tethered IP<sub>4</sub>, <sup>4</sup> hydrogen-labile benzyl protecting groups were employed at the C-2 and C-6 hydroxyls. Since base lability and ease of migration of the acetyl group can cause serious problems in polyhydroxy derivatives, benzylation at the C-2 and C-6 hydroxyls required mild, neutral conditions. Benzylation using the trichloroacetimidate method was unsuccessful due to the low solubility of the diol 7 and the acid sensitivity of the *p*-methoxybenzyl (PMB) groups. Instead, the more reactive benzyloxymethyl (BOM) group was selected. Although reaction of diol 7 with benzyloxymethyl chloride and diisopropylethylamine<sup>16</sup> failed, the BOM groups were readily introduced at C-2

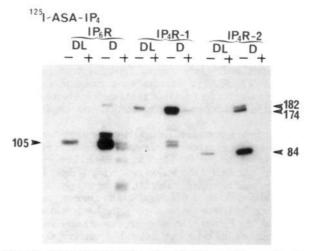


Figure 2. Photoaffinity labeling of purified  $IP_4$  and  $IP_6$  receptors by racemic (DL) and optically active (D)  $IP_4$  photolabel 13. Proteins purified from rat cerebellum by  $IP_4$  affinity chromatography<sup>6</sup> were photolabeled with 13 (10 nM) as described in the absence (–) or presence (+) of 200  $\mu$ M  $IP_4$ .<sup>7</sup> DL and D preparations of 13 were nominally of the same specific activity, based on synthesis using identical amounts and aliquots of the same batch of [125]NaI; equal molar equivalents were present in each incubation sample.  $IP_6R$  is the first eluting peak, and  $IP_4R$ -1 and  $IP_4R$ -2 are the second and third eluting peaks from the affinity column; the molecular sizes (in kDa) of the major bands are indicated.

and C-6 in 69% yield with BOM-Cl, H<sup>+</sup> sponge, and n-Bu<sub>4</sub>NBr in CH<sub>3</sub>CN.<sup>17</sup> To minimize acetyl migration, the BOM groups were introduced stepwise (23 °C for 10 h, 35 °C for 8 h, then 55 °C for 3 h). Basic methanolysis gave the C-1 alcohol 8 (95%), which served as the pivotal intermediate to a series of enantiomerically pure, P-1-modified IP<sub>4</sub> analogues.

Condensation of the alcohol 8 with (benzyloxy)[(3-(N-carbobenzoxyamino)propyl)oxy](diisopropylamino)phosphine<sup>5c</sup> in the presence of tetrazole and subsequent oxidation with MCPBA furnished the fully protected aminopropyl-tethered inositol 9 in 85% yield. Removal of the PMB groups with DDQ in wet CH<sub>2</sub>Cl<sub>2</sub> gave the triol 10 in 78% yield after SiO<sub>2</sub> chromatography. Phosphitylation of the triol followed by MCPBA oxidation (as for 8) gave the fully protected IP<sub>4</sub> derivative 11 (73% yield). Hydrogenolysis removed the benzyl and BOM groups to provide the optically active, P-1 aminopropyl tethered D-myo-Ins(1,3,4,5)P<sub>4</sub> (3) in quantitative yield after ion-exchange chromatography (Chelex, sodium form). The proton-decoupled <sup>31</sup>P NMR spectrum of this compound showed four singlets, δ 0.48, 3.59, 4.21, and 5.29 ppm, corresponding to the C-1 phosphodiester and the C-3, C-4, and C-5 phosphates, respectively.

Coupling 3 to N-hydroxysuccinimide-activated agarose (Affi-Gel 10) buffered at pH 8.5 provided an enantiomerically pure IP<sub>4</sub> affinity column, which has been employed in the purification of the IP<sub>4</sub>R (data not shown). Furthermore, reaction of this intermediate with N-hydroxysuccinimide 4-azidosalicylate in aqueous DMF, followed by radioiodination using Iodobeads, furnished the enantiomerically pure D-[125I]ASA-IP<sub>4</sub> photolabel 12. Figure 2 compares the efficacy of the optically active photolabel with that of the racemic material. It is evident from the intensity of the bands that D-12 is a substantially more sensitive and efficient probe than DL-12 for active-site modification of the IP<sub>n</sub> binding sites of these receptors.

Optically active derivatives of IP<sub>4</sub> modified at C-1 are now readily produced via a Ferrier rearrangement of glucose. Applications of this methodology in the synthesis of C-1 tritium labeled D-IP<sub>4</sub> and its photolabile derivatives for the characterization of the IP<sub>4</sub> binding site are in progress.

<sup>(5)</sup> For examples of P-1-tethered IP<sub>3</sub>, see: (a) Schäfer, R.; Nehls-Sahabandu, M.; Grabowski, B.; Dehlinger-Kremer, M.; Schultz, I.; Mayr, G. W. Biochem. J. 1990, 272, 817-825. (b) Lampe, D.; Potter, B. V. L. J. Chem. Soc., Chem. Commun. 1990, 1500-1501. (c) Prestwich, G. D.; Marecek, J. F.; Mourey, R. J.; Theibert, A. B.; Ferris, C. D.; Danoff, S. K.; Snyder, S. H. J. Am. Chem. Soc. 1991, 113, 1822-1825. (d) Prestwich, G. D.; Marecek, J. F. In Inositol Phosphates and Related Compounds: Synthesis and Therapeutic Potential; Reitz, A., Ed.; ACS Symposium Series 463; American Chemical Society: Washington, DC, 1991; Chapter 9, pp 122-131.

<sup>(6)</sup> Theibert, A. B.; Estevez, V. A.; Ferris, C. D.; Danoff, S. K.; Barrow, R. K.; Prestwich, G. D.; Snyder, S. H. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 3165–3169.

<sup>(7)</sup> Theibert, A. B.; Estevez, V. A.; Mourey, R. J.; Marecek, J. F.; Barrow, R. K.; Prestwich, G. D.; Snyder, S. H. J. Biol. Chem., in press.

<sup>(8)</sup> Challis, R. A. J.; Willcocks, A. L.; Mulloy, B.; Potter, B. V. L.; Nahorski, S. R. Biochem. J. 1991, 274, 861-867.

<sup>(9)</sup> A short, practical synthesis of optically active D-myo-IP<sub>4</sub> (2) has been reported: Watanabe, Y.; Fujimoto, T.; Shinohara, T.; Ozaki, S. J. Chem. Soc., Chem. Commun. 1991, 428-429.

<sup>(10)</sup> Bender, S. L.; Budhu, R. J. J. Am. Chem. Soc., preceding paper in this issue.

<sup>(11)</sup> For a synthesis of optically active myo-inositol from glucose, see: Jaramillo, C.; Martin-Lomas, M. Tetrahedron Lett. 1991, 32, 2501-2504.

<sup>(12)</sup> Chaudhary, S. K.; Hernandez, O. Tetrahedron Lett. 1979, 95-98.

<sup>(13) (</sup>a) Conventional detritylation with trifluoroacetic acid/CH<sub>2</sub>Cl<sub>2</sub> resulted in partial removal of the p-methoxybenzyl groups. (b) All compounds were fully characterized by <sup>1</sup>H, <sup>13</sup>C, and <sup>31</sup>P NMR, FAB-MS, and for new compounds, elemental analysis.

<sup>(14)</sup> Cook, S. L.; Secrist, J. A., III. J. Am. Chem. Soc. 1979, 101, 1554-1564.

<sup>(15)</sup> Evans, D. A.; Chapman, K. T.; Carreira, E. M. J. Am. Chem. Soc. 1988, 110, 3560-3578.

<sup>(16)</sup> Stork, G.; Isobe, M. J. Am. Chem. Soc. 1975, 97, 6260-6261.

<sup>(17)</sup> Evans, D. A.; Bender, S. L. Tetrahedron Lett. 1986, 27, 799-802.
(18) Theibert, A. B.; Estevez, V. A.; Barrow, R. K.; Prestwich, G. D.;
Snyder, S. H., unpublished results.

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Supplementary Material Available: Full experimental and spectral details for this synthesis (10 pages). Ordering information is given on any current masthead page.

# Catalytic Asymmetric Synthesis of Optically Active 2-Alkanols via Hydrosilylation of 1-Alkenes with a Chiral Monophosphine—Palladium Catalyst

Yasuhiro Uozumi and Tamio Hayashi\*

Catalysis Research Center and Graduate School of Pharmaceutical Sciences Hokkaido University, Kita-ku, Sapporo 060, Japan Received August 23, 1991

Catalytic asymmetric functionalization of simple prochiral olefins is an important goal in synthetic organic chemistry. We report here the first successful conversion of alkyl-substituted terminal olefins into optically active secondary alcohols (>94% ee), which is realized by palladium-catalyzed asymmetric hydrosilylation in the presence of a new chiral monodentate phosphine ligand (MOP, 1) followed by oxidation of the carbon-silicon bond (Scheme I).

It is well-documented<sup>5</sup> that the hydrosilylation of terminal olefins is catalyzed by platinum, rhodium, or nickel complexes to proceed with anti-Markovnikoff selectivity leading to 1-silylalkanes. Rather surprisingly, only a little attention has been paid to the use of palladium catalysts for the hydrosilylation of 1-alkenes<sup>6,7</sup> in spite of their frequent use for the reaction of 1,3-dienes

(2) (a) Zhang, W.; Loebach, J. L.; Wilson, S. R.; Jacobsen, E. N. J. Am. Chem. Soc. 1990, 1/2, 2801 and references cited therein. (b) Sharpless, K. B.; Amberg, W.; Beller, M.; Chen, H.; Hartung, J.; Kawanami, Y.; Lübben, D.; Manoury, E.; Ogino, Y.; Shibata, T.; Ukita, T. J. Org. Chem. 1991, 56, 4585 and references cited therein.

(3) Asymmetric synthesis of 1-arylethanols via palladium-catalyzed hydrosilylation or rhodium-catalyzed hydroboration of vinylarenes has been reported: (a) Hayashi, T.; Tamao, K.; Katsuro, Y.; Nakae, I.; Kumada, M. Tetrahedron Lett. 1980, 21, 1871. (b) Hayashi, T.; Matsumoto, Y.; Ito, Y. J. Am. Chem. Soc. 1989, 111, 3426. (c) Zhang, J.; Lou, B.; Guo, G.; Dai, L. J. Org. Chem. 1991, 56, 1670.

(4) (a) Tamao, K.; Ishida, N.; Tanaka, T.; Kumada, M. Organometallics 1983, 2, 1694. (b) Tamao, K.; Ishida, N. J. Organomet. Chem. 1984, 269, C37. (c) Tamao, K.; Nakajo, E.; Ito, Y. J. Org. Chem. 1987, 52, 4412. (d) Tamao, K. In Organosilicon and Bioorganosilicon Chemistry; Sakurai, H., Ed. Ellis Horwood: Chichester, 1985, p. 231

Ed.; Ellis Horwood: Chichester, 1985; p 231.

(5) For reviews: (a) Ojima, I. In The Chemistry of Organic Silicon Compounds; Patai, S., Rappoport, Z., Eds.; John Wiley: Chichester, 1989; p 1479. (b) Speier, J. L. Advanced Organometallic Chemistry; Stone, F. G. A., West, R., Eds.; Academic Press: New York, 1979; Vol. 17, p 407. (c) Lukevics, E.; Belyakova, Z. V.; Pomerantseva, M. G.; Voronkov, M. G. In Journal of Organometallic Chemistry Library; Seyferth, D., Davies, A. G., Fisher, E. O., Normant, J. F., Reutov, O. A., Eds.; Elsevier: Amsterdam, 1977; Vol. 5. (d) Ojima, I.; Kogure, T. Rev. Silicon, Germanium, Tin Lead Compd. 1981, 5, 7.

(6) It has been reported that reaction of 1-octene with HSiCl<sub>3</sub> catalyzed by Pd(PPh<sub>3</sub>)<sub>4</sub> at 100 °C gives 1-octylsilane in 85% yield: Tsuji, J.; Hara, M.; Ohno, K. Tetrahedron 1974, 30, 2143.

(7) Regioselective hydrosilylation of CF<sub>3</sub>CH=CH<sub>2</sub> with HSiMeCl<sub>2</sub> or HSiCl<sub>3</sub> catalyzed by PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> has been reported: Ojima, I.; Yatabe, M.; Fuchikami, T. J. Organomet. Chem. 1984, 260, 335.

### Scheme I

### Scheme IIa

(a: X = OMe, b: X = OPr-i, c:  $X = OCH_2Ph$ , d: X = Et)

 $^a$ (a) (i) 3 N NaOH, 1,4-dioxane, methanol; (ii) MeI, i-PrI or PhCH<sub>2</sub>Br (3-10 equiv),  $K_2$ CO<sub>3</sub> (2-4 molar equiv), acetone, reflux, 3-24 h (7a, 99%; 7b, 92%; 7c, 87%). (b) EtMgBr (1.1 equiv), NiCl<sub>2</sub>-(dppe) (2 mol %), Et<sub>2</sub>O, reflux, 24 h (7d, 81%). (c) Et<sub>3</sub>N (7-20 equiv), Cl<sub>3</sub>SiH (5 equiv), xylene, 120 °C, 3-5 h (1a, 97%; 1b, 84%; 1c, 96%; 1d, 79%).

and styrenes.<sup>5,6</sup> In order to develop a catalyst possessed of high catalytic activity, high regioselectivity giving 2-silylalkanes, and high enantioselectivity in addition, we examined several types of phosphine-palladium catalysts for the reaction of 1-hexene (2a) with trichlorosilane. It was found that palladium complexes coordinated with a chelating bis(phosphine), dppb,<sup>8</sup> chiraphos,<sup>9</sup> or BINAP,<sup>10</sup> did not catalyze the hydrosilylation at 80 °C, while the reaction took place at 40 °C with monodentate phosphine ligands.<sup>11,12</sup> Among the monodentate phosphine ligands, (S)-2-(diphenylphosphino)-2'-methoxy-1,1'-binaphthyl (MOP, 1a)<sup>13</sup> turned out to be by far the best ligand, giving a high yield of 2-(trichlorosilyl)hexane (3a) with high regioselectivity<sup>14</sup> as well as high enantioselectivity.

Chiral monophosphines (MOPs, 1) are the ligands we have designed with a view to using them for the catalytic asymmetric reactions where a monophosphine ligand is required to generate a catalytically active species.<sup>15</sup> The present palladium-catalyzed hydrosilylation is one of the cases. The phosphines 1a-d were readily prepared in high yields starting with known optically active

(8) 1,4-Bis(diphenylphosphino)butane.

(9) (S,S)-2,3-Bis(diphenylphosphino)butane: Fryzuk, M. D.; Bosnich, B. J. Am. Chem. Soc. 1977, 99, 6262.

(10) (R)-2,2'-Bis(diphenylphosphino)-1,1'-binaphthyl: Takaya, H.; Mashima, K.; Koyano, K.; Yagi, M.; Kumobayashi, H.; Taketomi, T.; Akutagawa, S.; Noyori, R. J. Org. Chem. 1986, 51, 629.

(11) For example, the reaction in the presence of 0.1 mol % of a palladium-triphenylphosphine catalyst (P/Pd = 2/1) at 40 °C for 24 h gave a 12% yield of hexylsilanes consisting of 1- and 2-isomers in a ratio of 91/9, accompanied by isomerization of 1-hexene into internal olefins. See also ref 6.

(12) It is reasonable to expect that a monodentate phosphine ligand generates a palladium catalyst that is more active for the hydrosilylation than a chelating bis(phosphine) ligand. The former can form square-planar palladium(II) intermediate  $PdH(SiCl_3)L(CH_2 - CHR)$  (L = monophosphine), which offers a coordination site for the activation of olefin.

(13) Preparation of (S)-1a from (S)-2'-methoxy-1,1'-binaphthyl-2-carboxylic acid has been reported: Hattori, T.; Shijo, M.; Kumagai, S.; Miyano, S. Chem. Express 1991, 6, 335.

(14) The high catalytic activity and regioselectivity of the palladium-MOP complex may be related to the reactivity of key intermediate Pd(2-alkyl)L-(silyl). It seems that MOP ligand can accelerate the reductive elimination of 2-silylalkane and/or retard the  $\beta$ -hydrogen elimination forming 2-alkenes. Triphenylphosphine or tri-o-tolylphosphine caused the isomerization of a terminal olefin into internal olefins during the hydrosilylation while MOP did not (see footnote 11).

(15) An example for those reactions is nickel-catalyzed asymmetric cross-coupling: Hayashi, T.; Hayashizaki, K.; Kiyoi, T.; Ito, Y. J. Am. Chem. Soc. 1988, 110, 8153.

<sup>(1)</sup> For recent reviews on catalytic asymmetric reactions: (a) Brunner, H. Synthesis 1988, 645. (b) Brunner, H. Top. Stereochem. 1988, 18, 129. (c) Consiglio, G.; Waymouth, R. M. Chem. Rev. 1989, 89, 257. (d) Noyori, R.; Kitamura, M. Modern Synthetic Methods; Scheffold, R., Ed.; Springer-Verlag: New York, 1989; Vol. 5, p 115. (e) Ojima, I.; Clos, N.; Bastos, C. Tetrahedron 1989, 45, 6901.